

Microbial Flora in Chronic Periodontitis: Study at a Tertiary Health Care Center from North Karnataka

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ABSTRACT

Background and Objective: Periodontitis is a major public health problem in India with a prevalence of 60–80%. If untreated it acts as a risk factor for systemic diseases. Data on anaerobic periodontal microflora in the Indian population is very scarce. Hence, this study was undertaken to know the nature of oral microbiota in chronic periodontitis in this region of India and also the semiquantitative study in pre- and post-treatment group and to determine antibiotic susceptibility pattern for aerobic isolates.

Materials and Methods: The present study was conducted on 60 cases. Material was collected from the subgingival pockets in patients with chronic periodontitis attending the Periodontology, Outpatient Department. Clinical samples were transported to the laboratory in fluid thioglycollate medium. Initially Gram's stain and Fontana stains were done. Aerobic, anaerobic, and microaerophilic culture were put up. Antibiotic sensitivity test was done for aerobic isolates.

Results: Sixty samples yielded 121 isolates of which 78.34% were polymicrobial, 11.66% were monomicrobial and oral commensals were grown in 10% cases. Out of 121 isolates 91.74% were anaerobic, 7.43% were aerobic and 0.83% were microaerophilic. *Fusobacterium* species was the most common isolate among anaerobes. Using "paired *t*-test" "*P*" value was significant indicating significant reduction in colony count after phase-I periodontal therapy.

Conclusion: This study has shown that anaerobic bacteria are important cause of chronic periodontitis, along with aerobes and microaerophilic organisms. *Fusobacterium spp*, *Bacteroides fragilis*, *Porphyromonas spp* and *Prevotella intermedia* are the most common anaerobic pathogens. Bacterial culture methods are still economical and gold standard.

Key words: Aerobe, anaerobe, oral microbial flora, periodontitis, subgingival plaque

INTRODUCTION

Symbiosis is an intimate association between the host and one or more colonizing bacteria. Human beings are colonized with an enormous number of bacteria; by-and-large skin, gut, urogenital tract and oral cavity are the typical sites. However, oral cavity is the one body site where co-operativity between the bacteria and host breaks down. If proper care is not

taken to remove the plaque bacteria, mouth is prone to very common diseases like caries and periodontitis.^[1]

Periodontitis is defined as an inflammatory disease of the supporting tissues of the teeth caused by specific microorganisms or group of specific microorganisms resulting in progressive destruction of periodontal ligament and alveolar bone with pocket formation, recession or both.^[2] The disease prevalence is 13–57% worldwide and 65–80% in India accounting to a major health problem.^[3,4]

Bacterial DNA sequencing study has suggested that nearly 19,000 bacterial phylotypes exist in the oral cavity. Nearly, 500 bacterial strains have been recovered from subgingival plaque, but only a small number are potential pathogens.^[5]

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Immunological, DNA probe assessment, bacterial enzyme detection and culture are the methods available for isolation of oral bacteria, among which culture remains economical and gold standard.^[6]

There are good number of studies available from western world regarding the microbiota of oral cavity both in normal and in periodontitis condition, but data on anaerobic periodontal microflora in the Indian population is very scarce. Hence, the study was undertaken to isolate and identify the predominant aerobic and anaerobic microflora associated with adult generalized periodontitis, semiquantitative study of isolated microflora of adult generalized periodontitis in pretreatment and posttreatment groups and to study the antibiotic sensitivity pattern of aerobic isolates in pretreatment group.

MATERIALS AND METHODS

This study was conducted in the Department of Microbiology at a Tertiary Care Teaching Hospital. Samples were collected from subgingival pockets in patients with chronic periodontitis attending the Periodontology Outpatient Department at our Institute of Dental Sciences, over a period of 1-year from January to December 2010. The study comprised of 60 cases (Sample size was calculated using formula $4pq/d^2$ where p is prevalence, q is $100-p$ and d is 10% of the error).

Clinically diagnosed all new cases of chronic periodontitis were included in the study and patients with history of systemic conditions such as diabetes mellitus, nutritional deficiencies, pregnant woman, antibiotic usage in the last 3 months and patients with history of undergoing any dental procedures in the last 3 months were excluded. Samples were collected twice that is, in pretreatment and post-treatment periods after phase I therapy (scaling and root planning [SRP] along with amoxicillin and metronidazole for 1-week).

Sample collection

Tooth surfaces were dried with sterile gauze to avoid contamination by saliva. Sub gingival plaque sample was collected from most pathological site using sterile periodontal Gracy curette and placed in fluid thioglycollate medium in a test tube and brought to the microbiology laboratory and processed immediately.

Laboratory methods

In the laboratory sample in fluid thioglycollate medium was transferred to 1 ml of trypticase soya broth and vortexing

was done for 1 min. Direct smears were prepared from the samples and Hucker's modification of Gram stain for anaerobes and modified Fontana staining methods were followed.^[7,8] With a standard loop, a loopful of sample was taken from vortexed solution and inoculated onto 5% sheep blood agar (SBA), Macconkey agar (MA), brain heart infusion agar and blood agar supplemented with haemin and Vitamin K, Kanamycin Vancomycin laked blood agar (KVLB) and Bacteroid Bile Esculin Agar (BBE). Lawn culture was done and the plates were incubated at 37°C for 24 h aerobically (MA, SBA), under 5% CO₂ (BA and BHI) and anaerobically (BBA supplemented with Vitamin K and hemin, KVLB, BBE). All aerobic isolates were identified and characterized biochemically by standard procedures as described by Mackie and McCartney practical medical microbiology.^[9]

Anaerobic organism isolation

Semi-quantitative estimation of organisms was done by counting the number of colonies and taking into consideration of significant number of colonies (>50 colonies) and then subculturing on BBA. From the purity BA plate colony morphology, pigment, hemolysis, fluorescence and pitting were recorded and Gram stain (modified for anaerobic), spot indole test and catalase tests were performed. Further identification was made using sodium polynethenol sulfonate disc, nitrate disc, kanamycin (1 mg), vancomycin (5 µg) and colistin (10 µg) discs. The method used for obtaining anaerobiosis in the jar was "internal gas generating system" described by Vaidhyalingam and Laxminarayana.^[10]

Antibiotic sensitivity testing

The antimicrobial susceptibility testing was done for aerobic isolates by disc diffusion method as described by Kirby and Bauer on Mueller-Hinton agar.^[9]

Statistical analysis

Statistical analysis was performed using "paired *t*-test" comparing two groups.

RESULTS

A total of 120 samples (60 pre-treatment and 60 post-treatment) were collected from 60 patients. Our cases ranged between 22 and 70 years, maximum number of cases (22) were in the age group of 38–47 years and least (1) in the age group of 68–77 years. Of 60 cases, 33 (55%) were female and 27 (45%) were male. Sixty samples yielded

121 isolates of which 47 (78.34%) were polymicrobial, 7 (11.66%) were monomicrobial and oral commensals were grown in 6 (10%) cases.

Out of 47 polymicrobial isolates 27 (57.45%) were combination of 2 isolates and 20 (42.55%) were combination of 3 isolates. Out of 121 isolates 111 (91.74%) were anaerobic, 9 (7.43%) were aerobic and 01 (0.83%) was microaerophilic.

Out of 111 anaerobic isolates 87 (78.4%) were Gram-negative bacilli (GNB) and 24 (21.6%) were Gram-positive cocci (GPC). Among GNB *Fusobacterium* species was the most common isolate [Figure 1]. Out of 24 GPC, *Micromonas micros* were the most common isolate 17 (70.83%), followed by *Peptostreptococcus anaerobius* 4 (16.67%), *Schleiferella asaccharolytica* 2 (8.33) and *Streptococcus mutans* 1 (4.17%).

Aerobic isolates were *Staphylococcus aureus*, *Streptococcus pneumoniae* and *Klebsiella pneumoniae* three each. The antibiotic susceptibility pattern of aerobic isolates is shown in Table 1. The only microaerophilic organism isolated was *Aggregatibacter actinomycetemcomitans* [Figure 2]. Spirochetes (*Treponema denticola*) were observed in 19 (31.66%) cases by modified Fontana staining from direct plaque sample smears. The result of semiquantitative study is shown in Table 2.

Using “paired *t*-test,” “*p*” value was significant indicating significant reduction in colony count after phase-I periodontal therapy. Paired *t* = 3.555, *P* = 0.001.

DISCUSSION

The search for the periodontal pathogens has been underway for more than 100 years and continues up today.

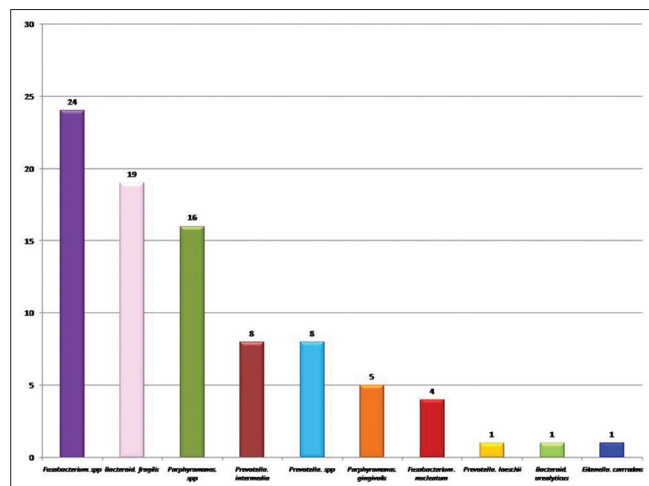


Figure 1: Anaerobic Gram-negative bacilli

The three factors which will determine the occurrence of active periodontal disease are a susceptible host, presence of pathogenic species and absence of so-called beneficial bacteria.^[11] The goal of the present study was to attempt to understand the nature of microbial complexes that exist in subgingival plaque.

In the present study, the cases ranged from 20 to 70 years with a maximum number of cases in the age group of 38–47 years. In contrast, Sixou *et al.*^[12] observed periodontitis cases in age group of 18–52 years, Antony *et al.*^[13] in the age group of 30–60 years, Kamma *et al.*^[14] in the age group of 14–35 years. It is a good old concept in the literature that the prevalence of periodontitis increases as age increases because of loss of periodontal tissue support, however maximum number of cases in our study were in adult group as observed by the previous authors.^[15]

The male to female ratio in our study was 0.8:1. That is, females were more affected than males as observed in studies by Sixou *et al.*,^[12] Salari and Kadkhoda^[16] Daniluk *et al.*,^[5] Nonnenmacher *et al.*^[17] and Socransky *et al.*^[6] In contrast in a study conducted by Sixou *et al.*^[18] the ratio was M:F::1.3:1. However, there is no established, inherent difference between men versus women in their susceptibility to periodontitis.^[15] 78.34% of our isolates were polymicrobial in nature in comparison to 11.06% of monomicrobial isolates. Our observations are consistent with the literature.^[4,12,16,17,19] Polymicrobial nature of periodontal plaque has been well established. In the studies conducted by Sixou *et al.*^[12] Nonnenmacher *et al.*^[17] Saini *et al.*^[4] and Mane *et al.*^[19] 97.14%, 100%, 72.28% and 97.3% of their isolates were polymicrobial, respectively. In contrast Salari and Kadkhoda^[16] observed 18.28% of their isolates were polymicrobial.



Figure 2: Low power microscopic picture of *Aggregatibacter actinomycetemcomitans* showing central star shaped colony

Table 1: Antibiotic susceptibility pattern of aerobic isolates

Isolates	Number of isolates	Antibiotic sensitivity											
		Cf		E		Ac		Ce		Ak		Ox	
		S	R	S	R	S	R	S	R	S	R	S	R
<i>Staphylococcus aureus</i>	03	3	0	3	0	3	0	3	0	NT	NT	3	0
<i>Streptococcus pneumoniae</i>	03	2	1	3	0	3	0	3	0	NT	NT	3	0
<i>Klebsiella pneumoniae</i>	03	2	1	NT	NT	3	0	3	0	3	0	NT	NT

S: Sensitive, R: Resistant, NT: Not tested, Cf: Ciprofloxacin, E: Erythromycin, Ac: Amoxiclav, Ak: Amikacin, Ox: Oxacillin, Ce: Cefotaxime

Table 2: Results of semiquantitative study

Pre-treatment (n=121)	Post-treatment (n=110)	Mean reduction (n=110)
750, 160±2, 267, 760=7.5×10 ⁵ ±2.2×10 ⁶	123.4±233.76	802, 530±2, 367, 570=8×10 ⁵ ±2.3×10 ⁶

Paired t=3.555, P=0.001

Out of 78.34% of polymicrobial isolates 57.44% were a combination of two isolates, and 42.55% were a combination of three isolates. In a study conducted by Saini *et al.*^[4] in India, they observed two isolates in 91.66% and three in 8.2%. In an another study conducted by Mane *et al.*^[19] in India observed two isolates in 8%, three and four isolates 40% each and five isolates in 12%.

The results of the present study show the diversity of anaerobic bacteria in chronic periodontitis. Anaerobic bacteria were isolated in 91.74% of periodontitis cases. Various studies from India and other countries showed an isolation rate of strict anaerobes ranging from 42% to 100% in periodontitis cases.^[4,16,19,20] In the two different studies conducted in France, detected 91.42% and 80.77% of anaerobic isolates. In a study by Mane *et al.*^[19] 83% were anaerobes. Whereas studies conducted by Salari and Kadkhoda,^[16] Daniluk *et al.*,^[5] Nonnenmacher *et al.*^[17] and Saini *et al.*^[4] detected comparatively lower number of anaerobes that is, 41.22%, 57.1%, 53.84% and 64.25%, respectively. The varying recovery rates of isolation can be due to varying criteria of patient selection, culture method employed, geographical differences, and molecular technique used for identification.^[4,19]

In the present study, Gram-negative anaerobes (78.4%) were predominantly isolated than the Gram-positive (21.6%) in the periodontitis cases. And our observations are comparable to the literature.^[4-6,16,19] *Fusobacterium spp* (27.60%), *Bacteroides fragilis* (21.84%), *Porphyromonas spp* (24.48%), *Prevotella intermedia* (9.20%) [Figure 3], *Fusobacterium nucleatum* (4.60%) and *Prevotella spp* (9.20%) were the most common organisms isolated in our study. Other workers have isolated similar anaerobes, but in varying proportions. In an Indian study by Saini *et al.*^[4] reported a much higher number of these isolates *Porphyromonas spp* 40%, *P. intermedia* 12%, *Prevotella* species 14%, *Fusobacterium nucleatum* 24% and *Bacteroides*

**Figure 3: Prevotella intermedia**

5%. Another Indian study by Mane *et al.*^[19] reported higher number of *P. intermedia* (17.6%), *F. nucleatum* (41%), *Fusobacterium spp* (29.41%) were in accordance with our results. Sixou *et al.*^[18] reported higher number of *P. intermedia* (15.78%) and *F. nucleatum* (7.01%). While Salari and Kadkhoda^[16] and Nonnenmacher *et al.*^[17] reported much lower number of these isolates. Comparison of spectrum of anaerobes in different studies is shown in Table 3. Beena *et al.*^[21] concluded from their study on periodontal pathogen that, as the infection progress the proportion of anaerobes especially GNB like *Prevotella*, *Porphyromonas* and *Fusobacterium* increases.

The Gram-positive anaerobic cocci constituted 21.6% of total anaerobic isolates in our study. Out of which *M. micros* (70.83%) were the most common isolates. Other GPC isolated in our study were *P. anaerobius* (16.67%), *S. asachrolytica* (8.33%) and *S. mutans* (4.17%).

Saini *et al.*^[4] and Nonnenmacher *et al.*^[17] reported *M. micros* in 23% and 51% respectively, which is much lower than our study. Sixou *et al.*^[12] observed 52.94% of *M. micros*, 5.88% of *P. anaerobius*. In another study by Sixou *et al.*,^[18] there were 14.28% each of *M. micros* and *P. anaerobius*. Salari and Kadkhoda^[16] reported 2.9% and van Winkelhoff *et al.*^[20] reported 70%, which is in accordance with our study. *M. micros* is considered to be a pathogen in the etiology of

Table 3: Comparison of spectrum of gram negative anaerobes in the literature

Gram-negative anaerobes	Mane et al. ^[19]	Salari and Kadkhoda ^[16]	Saini et al. ^[4]	Socransky et al. ^[6]	Nonnenmacher et al. ^[17]	Present study %
<i>Fusobacterium spp.</i>	-	-	29.4	7	-	27.60
<i>Bacteroides fragilis</i>	-	-	-	-	-	21.84
<i>Porphyromonas spp.</i>	48	24.5	14	5.26	6.4	24.48
<i>Prevotella intermedia</i>	12	7	17.4	15.78	10.2	9.2
<i>Prevotella species</i>	14	5	6	-	-	9.2
<i>Fusobacterium nucleatum</i>	24	-	41	7.01	1.3	4.6
<i>Prevotella loeschii</i>	-	-	-	-	-	1.14
<i>Bacteroides ureolyticus</i>	05	-	-	3.10	-	1.14
<i>Eikenella corrodens</i>	-	22	-	-	-	1.14
<i>Veillonella spp.</i>	09	-	-	-	-	-

mixed anaerobic infections more often and in increased percentage from patients with periodontitis, especially in subjects with active disease.^[22-26] Our results are in accordance with the previous observation suggesting that *M. micros* may be associated with periodontitis.

In the present study group, aerobes constituted 7.43%, while Daniluk et al.^[5] and Mane et al.^[19] isolated 42.9% and 14% of aerobes, respectively. The aerobes isolated were *S. aureus* (33.33%), *S. pneumoniae* (33.33%) and *K. pneumoniae* (33.33%) and our results are in accordance with studies of Daniluk et al.^[5] who reported *Staphylococcus* species (16.66%) and *Streptococcus* species (38.88%). Saini et al.^[4] reported 42.85% of *S. aureus* and 44.44% of *Klebsiella spp.* They also reported *Escherichia coli*, *Pseudomonas spp* (14.28%) and nonfermenters (28.5%), which are not observed in our study.^[4,5]

The only microaerophilic bacteria isolated in the present study is *A. actinomycetemcomitans* which accounts for 0.83% of total bacterial isolates. The isolation rate of *A. actinomycetemcomitans* from adult periodontitis was reported to be 53% by Antony et al.,^[13] 9% by Nonnenmacher et al.,^[17] 20.6% by Salari and Kadkhoda^[16] and 14.6% by Saini et al.^[4] These results show that *A. actinomycetemcomitans* seems to play a significant role in periodontitis along with the other anaerobes.

Spirochetes such as *T. denticola* were observed by modified Fontana staining in 31.66% of cases. In this study, there was significant decrease in bacterial count following SRP with $P < 0.001$, as observed in the literature. Two systemic reviews have suggested that adjuvant systemic antibiotic administration will have a better outcome than SRP alone.^[27,28] The predisposing factors for periodontitis include increasing age, prior history of periodontitis, smoking, stress, few drugs such as anticonvulsants, calcium channel blockers and oral contraceptive pills, systemic factors like HIV, Diabetes mellitus and hematological factors.^[29-31]

Periodontitis if untreated thought to act as a risk factor for systemic diseases like atherosclerosis, cardiovascular and cerebrovascular diseases, pregnancy complications, diabetes mellitus and others.^[31,32]

Since the concept of dental infection has been bacteriologically nonspecific and offers no rationale for antimicrobial treatment, careful assessment and isolation of both aerobes and anaerobes is the utmost importance in the treatment of orodental infection.^[4]

CONCLUSION

This study has shown that anaerobic bacteria are important cause of chronic periodontitis, along with aerobes and microaerophilic organisms. Cultural methods are still economical and gold standard. Considering the scarce data on microbial flora in the Indian population, further studies for assessment of microbial profile in various forms of periodontitis should be carried out.

REFERENCES

1. Nibali L, Donos N, Henderson B. Periodontal infectogenomics. J Med Microbiol 2009;58:1269-74.
2. Novak MJ. Classification of diseases and conditions affecting the periodontium. In: Newman MG, Takei HH, Klokkevold PR, Carranza FA, editors. Carranza's Clinical Periodontology. 10th ed. St. Louis Missouri: Elsevier; 2007. p. 103-4.
3. Rylev M, Kilian M. Prevalence and distribution of principal periodontal pathogens worldwide. J Clin Periodontol 2008;35:346-61.
4. Saini S, Aparna, Gupta N, Mahajan A, Arora DR. Microbial flora in orodental infections. Indian J Med Microbiol 2003;21:111-4.
5. Daniluk T, Tokajuk G, Cylwik-Rokicka D, Rozkiewicz D, Zaremba ML, Stokowska W. Aerobic and anaerobic bacteria in subgingival and supragingival plaques of adult patients with periodontal disease. Adv Med Sci 2006;51 Suppl 1:81-5.
6. Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent RL Jr. Microbial complexes in subgingival plaque. J Clin Periodontol 1998;25:134-44.
7. Forbes BA, Sahm DF, Wiessfeld AS. Bailey and Scott's Diagnostic Microbiology. 11th ed. St. Louis, Missouri: Mosby, Inc. An Affiliate of Elsevier Science; 2002.
8. Gangadhar NL, Rajashekhar M. A modified silver impregnation staining for leptospire. Indian Vet J 1998;75:349-51.

9. Collee JG, Fraser AG, Marmion BP, Simmons A. Mackie and McCartney Practical Medical Microbiology. 14th ed. Churchill Livingstone: Elsevier; 2008.
10. Vaidhyalingam K, Laxminarayana CS. Internal gas generator system suitable for creating anaerobiosis. Indian J Surg 1980;42:154-9.
11. Kesic L, Milasin J, Ijic M, Obradovic R. Microbial etiology of periodontal disease – Mini review. Med Biol 2008;15:1-6.
12. Sixou JL, Magaud C, Jolivet-Gougeon A, Cormier M, Bonnaure-Mallet M. Microbiology of mandibular third molar pericoronitis: Incidence of beta-lactamase-producing bacteria. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2003;95:655-9.
13. Antony B, Kurian R, Faizal M, Verma BR, Shivananda PG. Actinobacillus actinomycetemcomitans and anaerobes in periodontitis. Indian J Med Microbiol 1997;15:73-6.
14. Kamma JJ, Nakou M, Manti FA. Predominant microflora of severe, moderate and minimal periodontal lesions in young adults with rapidly progressive periodontitis. J Periodontol Res 1995;30:66-72.
15. Kinane DF, Lindhe J, Trombelli L. Chronic periodontitis. Lindhe J, Lang NP, Karring T, editors. Clinical Periodontology and Implant Dentistry. 5th ed., Vol. 1. Oxford: Blackwell Publishers; 2008. p. 398-495.
16. Salari MH, Kadkhoda Z. Rate of cultivable subgingival periodontopathogenic bacteria in chronic periodontitis. J Oral Sci 2004;46:157-61.
17. Nonnenmacher C, Mutters R, de Jacoby LF. Microbiological characteristics of subgingival microbiota in adult periodontitis, localized juvenile periodontitis and rapidly progressive periodontitis subjects. Clin Microbiol Infect 2001;7:213-7.
18. Sixou JL, Magaud C, Jolivet-Gougeon A, Cormier M, Bonnaure-Mallet M. Evaluation of the mandibular third molar pericoronitis flora and its susceptibility to different antibiotics prescribed in France. J Clin Microbiol 2003;41:5794-7.
19. Mane AK, Karmakar AP, Bharadwaj RS, Van Winkelhoff AJ, Loss BG, Van der Reijden WA, et al. Porphyromonas gingivalis, Bacteroids forsythus and other periodontal pathogens in subjects with and without periodontal destruction. J Clin Periodontol 2002;29:1023.
20. Van Winkelhoff AJ, Loss BG, Van der Reijden WA, Van der Velden U. Porphyromonas gingivalis, Bacteroids forsythus and other periodontal pathogens in subjects with and without periodontal destruction. J Clin Periodontol 2002;29:1023.
21. Beena VK, Francis J, Bhat M, Kotion M, Shivananda PG. Anaerobic bacteria in periodontal infections. J Indian Dent Assoc 1992;63:215-9.
22. Socransky SS, Haffajee AD. Microbial etiological agents of destructive periodontal diseases. In: Socransky SS, Haffajee AD, editor. Microbiology and Immunology of Periodontal Diseases. Oxford: Munksgaard, Blackwell; 1994. p. 78-111.
23. Murdoch DA, Mitchelmore IJ, Tabaqchali S. Peptostreptococcus micros in polymicrobial abscesses. Lancet 1988;1:594.
24. Moore WE, Moore LH, Ranney RR, Smibert RM, Burmeister JA, Schenkein HA. The microflora of periodontal sites showing active destructive progression. J Clin Periodontol 1991;18:729-39.
25. Rams TE, Feik D, Listgarten MA, Slots J. Peptostreptococcus micros in human periodontitis. Oral Microbiol Immunol 1992;7:1-6.
26. Tanner A, Kent R, Maiden MF, Taubman MA. Clinical, microbiological and immunological profile of healthy, gingivitis and putative active periodontal subjects. J Periodontol Res 1996;31:195-204.
27. Haffajee AD, Socransky SS, Gunsolley JC. Systemic anti-infective periodontal therapy. A systematic review. Ann Periodontol 2003;8:115-81.
28. Herrera D, Sanz M, Jepsen S, Needleman I, Roldán S. A systematic review on the effect of systemic antimicrobials as an adjunct to scaling and root planing in periodontitis patients. J Clin Periodontol 2002;29 Suppl 3:136-59.
29. Novak MJ, Novak KF. Chronic periodontitis. In: Newuman MG, Takei HH, Klokkevold PR, Carranza FA, editors. Carranza's Clinical Periodontology. 10th ed, St. Louis Missouri: Elsevier; 2007. p. 497-99.
30. Kinane DF, Lindhe J, Trombelli L. Chronic periodontitis. In: Lindhe J, Lang NP, editors. Clinical Periodontology and Implant Dentistry. 5th ed. Oxford: Blackwell Publishers; 2008. p. 424-6.
31. Parameter on periodontitis associated with systemic conditions. American Academy of Periodontology. J Periodontol 2000;71 5 :876-9.
32. Williams RC, Paquette DW. Periodontal disease as a risk for systemic disease. In: Lindhe J, Lang NP, Karring T, editors. Clinical Periodontology and Implant Dentistry. 5th ed., Vol. 1. Oxford: Blackwell Publishers; 2008. p. 398-495.

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