

Supplementary Table 1 Primers used in this study

PCR	Target	Primer sequence (5'–3')	Amplicon size (bp)	Reference
I	<i>BimA_{Bm}</i> ^a	F: 5'-CGATACAGCCGACACAAGT-3' R: 5'-TGTTTTGACCGGACGCATTT-3'	240	
II	<i>BimA_{Bp}</i>	F: 5'-GGAAGCTTTGGCGTGCATAT-3' R: 5'-CCCATGCCTTCCTCGACTAAT-3'	60	2
III	<i>fhaB3</i>	F: 5'-GACGCGGCACGTCTGATC-3' R: 5'-CGCGGATAAACTCGGATTG-3'	58	
	LPS A	F: 5'-TCAAACCTATCCGCGTGTCCAAGT-3' R: 5'-TCGTCGTCAAGAAATCCCAGCCAT-3'	195	
	LPS B	F: 5'-AATCTTTTTCTGATTCCGTCG-3' R: 5'-ACCAGAAGACAAGGAGAAAGGCCA-3'	93	
	LPS B2	F: 5'-AACCGGGTAGTTCGCGATTAC-3' R: 5'-ATACGCCGGTGTAGAACAGTA-3'	364	

Abbreviations: LPS, lipopolysaccharide; PCR, polymerase chain reaction.

^aPrimers were designed by Dr. Tushar Shaw and Dr. Chiranjay Mukhopadhyay, Department of Microbiology, KMC Manipal, India.

Supplementary Table 2 Cycling conditions used in this study

PCR	Reaction volume	Primer concentration	PCR conditions	DNA template
I and II	25 µL	0.32 µM	95°C for 10 min 35 cycles 95°C for 30 s 60.9°C for 30 s 72°C for 30 s 72°C for 7 min	2 µL
III	25 µL	0.4 µM	95°C for 10 min 35 cycles 95°C for 30 s 59°C for 30 s 72°C for 30 s 72°C for 7 min	

Abbreviation: PCR, polymerase chain reaction.